

New Hampshire Volunteer Lake Assessment Program

2003 Interim Report for Forest Lake Winchester



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Water Division
Watershed Management Bureau
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OBSERVATIONS & RECOMMENDATIONS

After reviewing data collected from **FOREST LAKE, WINCHESTER**, the program coordinators have made the following observations and recommendations:

As you are aware, the Franklin Pierce College (FPC) satellite VLAP laboratory was not able to analyze samples during the 2003 sampling season. This was largely due to personnel and budget issues at the college. Although the FPC laboratory was not able to analyze samples, staff at FPC continued to lend out sampling equipment to volunteer monitors in this area. This was truly a cooperative effort between DES, FPC, and the volunteer monitors in this region. We want to thank you again for bearing with us this season. Also, we want to assure you that DES and FPC are working together to get the FPC lab up and running for the 2004 sampling season. We will keep you posted on the status of the laboratory as the sampling season approaches.

Thank you for your hard work sampling the lake/pond this season! Your monitoring group sampled **three** times this season! As you know, with multiple sampling events each season, we will be able to more accurately detect changes in water quality. Keep up the good work!

FIGURE INTERPRETATION

- **Figure 1 and Table 1:** The graphs in Figure 1 (Appendix A) show the historical and current year chlorophyll-a concentration in the water column. Table 1 (Appendix B) lists the maximum, minimum, and mean concentration for each sampling season that the lake/pond has been monitored through the program.

Chlorophyll-a, a pigment naturally found in plants, is an indicator of the algal abundance. Because algae are usually microscopic plants that contain chlorophyll-a, and are naturally found in lake ecosystems, the chlorophyll-a concentration measured in the water gives an estimation of the algal concentration or lake productivity.

The mean (average) summer chlorophyll-a concentration for New Hampshire's lakes and ponds is 7.02 mg/m³.

The current year data (the top graph) show that the chlorophyll-a concentration **decreased** from June to July, and then **remained relatively stable** from July to August. The chlorophyll-a concentration in June was **slightly less than** the state mean, while the concentration in July and August was **less than** the state mean.

The historical data (the bottom graph) show that the 2003 chlorophyll-a mean is **much less than** the state mean. In addition, it is important to point out the mean annual chlorophyll concentration this season (3.70 mg/m³) is the **lowest** annual mean since monitoring began in 1991.

Overall, visual inspection of the historical data trend line (the bottom graph) shows a **variable** in-lake chlorophyll-a trend. This means that the concentration has **fluctuated**, but has not *continually increased* or *continually decreased*, since monitoring began. In the 2004 annual report, we will conduct a statistical analysis of the historic data to objectively determine if there has been a significant change in the annual mean chlorophyll-a concentration since monitoring began.

While algae are naturally present in all lakes/ponds, an excessive or increasing amount of any type is not welcomed. In freshwater lakes/ponds, phosphorus is the nutrient that algae depend upon for growth. Algal concentrations may increase with an increase in nonpoint sources of phosphorus loading from the watershed, or in-lake sources of phosphorus loading (such as phosphorus releases from the sediments). Therefore, it is extremely important for volunteer monitors to continually educate residents about how activities within the watershed can affect phosphorus loading and lake/pond quality.

- **Figure 2 and Table 3:** The graphs in Figure 2 (Appendix A) show historical and current year data for lake/pond transparency. Table 3 (Appendix B) lists the maximum, minimum and mean transparency data for each sampling season that the lake/pond has been monitored through the program.

Volunteer monitors use the Secchi-disk, a 20 cm disk with alternating black and white quadrants, to measure water clarity (how far a person can see into the water). Transparency, a measure of water clarity, can be affected by the amount of algae and sediment from erosion, as well as the natural colors of the water. **The mean**

(average) summer transparency for New Hampshire's lakes and ponds is 3.7 meters.

The current year data (the top graph) show that the in-lake transparency **increased** from June to July, and then **decreased** from July to August. The transparency in June was **slightly less than** the state mean, in July was **slightly greater than** the state mean, and in August was **less than** the state mean.

The historical data (the bottom graph) show that the 2003 mean transparency is **slightly less than** the state mean.

Overall, visual inspection of the historical data trend line (the bottom graph) shows a **relatively stable** trend for in-lake transparency. Specifically, the transparency has **remained approximately the same (ranging between approximately 2.5 and 3.7 meters)** since monitoring began in 1991. As discussed previously, in the 2004 annual report, we will be able to conduct a statistical analysis of the historic data to objectively determine if there has been a significant change in the annual mean transparency since monitoring began.

Typically, high intensity rainfall causes erosion of sediments into lakes/ponds and streams, thus decreasing clarity. Efforts should continually be made to stabilize stream banks, lake/pond shorelines, disturbed soils within the watershed, and especially dirt roads located immediately adjacent to the edge of tributaries and the lake/pond. Guides to Best Management Practices designed to reduce, and possibly even eliminate, nonpoint source pollutants, such as sediment loading, are available from DES upon request.

- **Figure 3 and Table 8:** The graphs in Figure 3 (Appendix A) show the amounts of phosphorus in the epilimnion (the upper layer) and the hypolimnion (the lower layer); the inset graphs show current year data. Table 8 (Appendix B) lists the annual maximum, minimum, and median concentration for each deep spot layer and each tributary since the lake/pond has joined the program.

Phosphorus is the limiting nutrient for plant and algae growth in New Hampshire's freshwater lakes and ponds. Too much phosphorus in a lake/pond can lead to increases in plant and algal growth over time. **The median summer total phosphorus concentration in the epilimnion (upper layer) of New Hampshire's lakes and ponds is 11 ug/L. The median summer phosphorus concentration in the hypolimnion (lower layer) is 14 ug/L.**

The current year data for the epilimnion (the top inset graph) and the hypolimnion (the bottom inset graph) show that the phosphorus concentration **decreased** from June to July, and then **increased** from July to August. The phosphorus concentration in June was **slightly less than** the state median, in July was **less than** the state median, and in August was **greater than** the state median.

The historical data show that the 2003 mean epilimnetic and hypolimnetic phosphorus concentrations are **approximately equal to** the respective state medians.

Overall, visual inspection of the historical data trend line for the epilimnion and the hypolimnion shows a **variable** phosphorus trend. This means that the phosphorus concentration in the epilimnion and in the hypolimnion has **fluctuated**, but has not *continually increased* or *continually decreased* since monitoring began.

One of the most important approaches to reducing phosphorus loading to a waterbody is to continually educate watershed residents about its sources and how excessive amounts can adversely impact the ecology and value of lakes and ponds. Phosphorus sources within a lake or pond's watershed typically include septic systems, animal waste, lawn fertilizer, road and construction erosion, and natural wetlands.

TABLE INTERPRETATION

➤ **Table 2: Phytoplankton**

Table 2 (Appendix B) lists the current and historic phytoplankton species observed in the lake/pond. The dominant phytoplankton species observed this year were ***Asterionella* (a diatom), *Dinobryon* (a golden-brown algae), and *Cyclotella* (a diatom).**

Phytoplankton populations undergo a natural succession during the growing season (Please refer to the "Biological Monitoring Parameters" section of this report for a more detailed explanation regarding seasonal plankton succession). Diatoms and golden-brown algae are typical in New Hampshire's less productive lakes and ponds.

➤ **Table 2: Cyanobacteria (Blue-green algae)**

Small amounts of the cyanobacterium ***Anabaena* and *Microcystis*** were observed in the plankton sample this season. ***These species, if present in large amounts, can be toxic to livestock, wildlife, pets, and humans.***

Cyanobacteria can reach nuisance levels when excessive nutrients and favorable environmental conditions occur. During September of 2003, a few lakes and ponds in the southern portion of the state experienced cyanobacteria blooms. This was likely due to nutrient loading to these waterbodies. As mentioned previously, many weeks during the Spring and Summer of 2003 were rainy, which likely resulted in a large amount of nutrient loading to surface waters.

The presence of cyanobacteria serves as a reminder of the lake's/pond's delicate balance. Watershed residents should continue to act proactively to reduce nutrient loading into the lake/pond by eliminating fertilizer use on lawns, keeping the lake/pond shoreline natural, re-vegetating cleared areas within the watershed, and properly maintaining septic systems and roads.

In addition, residents should also observe the lake/pond in September and October during the time of fall turnover (lake mixing) to document any algal blooms that may occur. Cyanobacteria (blue-green algae) have the ability to regulate their depth in the water column by producing or releasing gas from vesicles. However, occasionally lake mixing can affect their buoyancy and cause them to rise to the surface and bloom. Wind and currents tend to "pile" cyanobacteria into scums that accumulate in one section of the lake/pond. If a fall bloom occurs, please contact the VLAP Coordinator.

➤ **Table 4: pH**

Table 4 (Appendix B) presents the in-lake and tributary current year and historical pH data.

pH is measured on a logarithmic scale of 0 (acidic) to 14 (basic). pH is important to the survival and reproduction of fish and other aquatic life. A pH below 5.5 severely limits the growth and reproduction of fish. A pH between 6.5 and 7.0 is ideal for fish. The mean pH value for the epilimnion (upper layer) in New Hampshire's lakes and ponds is **6.5**, which indicates that the surface waters in state are slightly acidic. For a more detailed explanation regarding pH, please refer to the "Chemical Monitoring Parameters" section of this report.

The mean pH at the deep spot this season ranged from **6.44** in the hypolimnion to **6.72** in the epilimnion, which means that the water is ***slightly acidic***. When organic matter is decomposed near the lake bottom, acidic by-products are produced which likely explains the lower pH (meaning higher acidity) in the hypolimnion.

Due to the presence of granite bedrock in the state and the deposition of acid rain, there is not much that can be done to effectively increase lake/pond pH.

➤ **Table 5: Acid Neutralizing Capacity**

Table 5 (Appendix B) presents the current year and historic epilimnetic ANC for each year the lake/pond has been monitored through VLAP.

Buffering capacity or ANC describes the ability of a solution to resist changes in pH by neutralizing the acidic input to the lake. The mean ANC value for New Hampshire's lakes and ponds is **6.7 mg/L**, which indicates that many lakes and ponds in the state are "highly sensitive" to acidic inputs. For a more detailed explanation, please refer to the "Chemical Monitoring Parameters" section of this report.

The mean Acid Neutralizing Capacity (ANC) of the epilimnion (the upper layer) this season was **7.33 mg/L**. Specifically, this indicates that the lake/pond is **highly sensitive** to acidic inputs (such as acid precipitation).

➤ **Table 6: Conductivity**

Table 6 (Appendix B) presents the current and historic conductivity values for tributaries and in-lake data. Conductivity is the numerical expression of the ability of water to carry an electric current. The mean conductivity value for New Hampshire's lakes and ponds is **62.1 uMhos/cm**. For a more detailed explanation, please refer to the "Chemical Monitoring Parameters" section of this report.

The in-lake conductivity has fluctuated, but, overall, has **slightly decreased** (meaning slightly improved) at the deep spot since monitoring began in 1991. In addition, the conductivity in the **Campground Inlet** has **decreased greatly**, and the conductivity in the Dump Branch Inlet has **decreased slightly** since monitoring began. This **decreasing** trend in conductivity suggests a reduction of pollutants in the watershed. We hope that this trend continues!

➤ **Table 8: Total Phosphorus**

Table 8 (Appendix B) presents the current year and historic total phosphorus data for in-lake and tributary stations. Phosphorus is the nutrient that limits the algae's ability to grow and reproduce. Please refer to the "Chemical Monitoring Parameters" section of this report for a more detailed explanation.

On the **July** sampling event, the total phosphorus concentration was **elevated** in the **Campground Inlet** (38 ug/L) and **NE Branch** (53 ug/L) samples. These stations have had a history of **fluctuating** total phosphorus concentrations. We recommend that your monitoring group conduct stream surveys and storm event sampling along these inlets so that we can determine what may be causing the elevated levels.

In addition, the phosphorus concentration in the **Dump Branch** (39 ug/L) was **elevated** on the August sampling event. The turbidity of the sample was also **elevated** (6.81 NTUs) and the laboratory staff indicated that organic matter was in the sample bottle. This suggests that the stream bottom may have been disturbed while sampling or that erosion is occurring in this portion of the watershed.

When the stream bottom is disturbed, sediment, which typically contains attached phosphorus, is released into the water column. When collecting samples in the inlets, please be sure to sample where there the stream is flowing and where the stream is deep enough to collect a “clean” sample.

If you suspect that erosion is occurring in this portion of the watershed, we recommend that your monitoring group conduct stream surveys and storm event sampling along this inlet. This additional sampling may allow us to determine what is causing the elevated levels of turbidity.

For a detailed explanation on how to conduct storm and stream surveys, please refer to the 2002 VLAP Annual Report “Special Topic Article”, or contact the VLAP Coordinator.

➤ **Table 9 and Table 10: Dissolved Oxygen and Temperature Data**

Table 9 (Appendix B) shows the dissolved oxygen/temperature profile(s) for the 2003 sampling season. Table 10 (Appendix B) shows the historical and current year dissolved oxygen concentration in the hypolimnion (lower layer). The presence of dissolved oxygen is vital to fish and amphibians in the water column and also to bottom-dwelling organisms. Please refer to the “Chemical Monitoring Parameters” section of this report for a more detailed explanation.

The dissolved oxygen concentration was **relatively high** at all depths sampled at the deep spot of the lake/pond on the June sampling event. However, as the summer progresses, oxygen typically becomes **depleted** in the hypolimnion (lower layer) of the pond by the process of decomposition. Specifically, the loss of oxygen in the hypolimnion results primarily from the process of biological oxidation of organic

matter (i.e.; biological organisms using oxygen to break down organic matter), both in the water column and particularly at the bottom of the lake/pond where the water meets the sediment. When oxygen levels are depleted to less than 1 mg/L in the hypolimnion (**as it has been in many past seasons**), the phosphorus that is normally bound up in the sediment may be re-released into the water column.

➤ **Table 11: Turbidity**

Table 11 (Appendix B) lists the current year and historic data for in-lake and tributary turbidity. Turbidity in the water is caused by suspended matter, such as clay, silt, and algae. Water clarity is strongly influenced by turbidity. Please refer to the “Other Monitoring Parameters” section of this report for a more detailed explanation.

As discussed previously, the turbidity in the **Dump Branch** inlet was **elevated** on the **August** sampling event, which suggests that the stream bottom may have been disturbed while sampling or that erosion is occurring in this portion of the watershed.

➤ **Table 12: Bacteria (*E.coli*)**

Table 12 lists the current year data for bacteria (*E.coli*) testing. *E. coli* is a normal bacterium found in the large intestine of humans and other warm-blooded animals. *E.coli* is used as an indicator organism because it is easily cultured and its presence in the water, in defined amounts, indicates that sewage **MAY** be present. If sewage is present in the water, potentially harmful disease-causing organisms may also be present.

E.coli sampling was conducted in multiple locations in the **Campground Inlet** on the July and August sampling events this season. While the results were **slightly elevated** (the highest result was 80 counts per 100 mL), the concentrations **did not exceed** the state standard of 88 counts per 100 mL for designated swimming beaches or the state standard of 406 counts per 100 mL for recreational waters that are not designated beaches.

We recommend that your group continue *E.coli* sampling along this inlet next season. If the results continue to be elevated, we will recommend that your group conduct a series of tests on a weekend during heavy beach use and also after a rain event. This additional sampling may help us determine the source of the bacteria.

For a detailed explanation on how to conduct rain event sampling, please refer to the 2002 VLAP Annual Report “Special Topic Article”, or contact the VLAP Coordinator.

DATA QUALITY ASSURANCE AND CONTROL

Annual Assessment Audit:

During the annual visit to your lake/pond, the biologist conducted a “Sampling Procedures Assessment Audit” for your monitoring group. Specifically, the biologist observed the performance of your monitoring group while sampling and filled out an assessment audit sheet to document the ability of the volunteer monitors to follow the proper field sampling procedures (as outlined in the VLAP Monitor’s Field Manual). This assessment is used to identify any aspects of sample collection in which volunteer monitors are not following the proper procedures, and also provides an opportunity for the biologist to retrain the volunteer monitors as necessary. This will ultimately ensure that the samples that the volunteer monitors collect are truly representative of actual lake and tributary conditions.

Overall, your monitoring group did an **excellent** job collecting samples on the annual biologist visit this season! Specifically, the members of your monitoring group followed the proper field sampling procedures. Keep up the good work!

Sample Receipt Checklist:

Each time your monitoring group dropped off samples at the laboratory this summer, the laboratory staff completed a sample receipt checklist to assess and document if the volunteer monitors followed proper sampling techniques when collecting the samples. The purpose of the sample receipt checklist is to minimize, and hopefully eliminate, future re-occurrences of improper sampling techniques.

Overall, the sample receipt checklist showed that your monitoring group did a **very good** job when collecting samples this season! Specifically, the members of your monitoring group followed the majority of the proper field sampling procedures when collecting and submitting samples to the laboratory. However, the laboratory did identify one aspect of sample collection that the volunteer monitors could improve upon.

- **Tributary Sampling:** Sediment/debris was observed in the white sample bottle for **Dump Branch** on the **August** sampling event. Please do not sample tributaries that are too shallow to collect a “clean” sample and do not sample the stream if the stream bottom has been disturbed. You may need to move upstream or downstream to collect a “clean” sample. If you disturb the stream bottom while

sampling, please rinse out the bottle and move to an upstream location and sample in an undisturbed area.

NOTES

- **Monitor's Note (6/10/03):** Sampling conducted after approx. 2 weeks of overcast and rainy weather. Forest Lake Campground sediment deposit into Campground Inlet. Newly exposed soil; sediment delta along shore and into lake. Land clearing upstream. Well-established milfoil in channel.
- (7/17/03):** Abundant milfoil observed.
- **Biologist's Note (7/17/03):** Phosphorous level at NE Branch Downstream and Campground Inlet was found to be elevated.
- (8/21/03):** The phosphorous levels at NE Branch Dump Branch were elevated.

USEFUL RESOURCES

Acid Deposition Impacting New Hampshire's Ecosystems, ARD-32, NHDES Fact Sheet, (603) 271-3505, or www.des.state.nh.us/factsheets/ard/ard-32.htm.

Best Management Practices to Control Nonpoint Source Pollution: A Guide for Citizens and Town Officials, NHDES-WD 97-8, NHDES Booklet, (603) 271-3503.

Camp Road Maintenance Manual: A Guide for Landowners. Kennebec Soil and Water Conservation District, 1992, (207) 287-3901.

Cyanobacteria in New Hampshire Waters Potential Dangers of Blue-Green Algae Blooms, NHDES Fact Sheet, (603) 271-3505, or www.des.state.nh.us/factsheets/wmb/wmb-10.htm.

Erosion Control for Construction in the Protected Shoreland Buffer Zone, WD-SP-1, NHDES Fact Sheet, (603) 271-3503 or www.des.state.nh.us/factsheets/sp/sp-1.htm

Impacts of Development Upon Stormwater Runoff, WD-WQE-7, NHDES Fact Sheet, (603) 271-3503, or www.des.state.nh.us/factsheets/wqe/wqe-7.htm

Lake Protection Tips: Some Do's and Don'ts for Maintaining Healthy Lakes, WD-BB-9, NHDES Fact Sheet, (603) 271-3503 or www.des.state.nh.us/factsheets/bb/bb-9.htm.

OBSERVATIONS AND RECOMMENDATIONS

2003

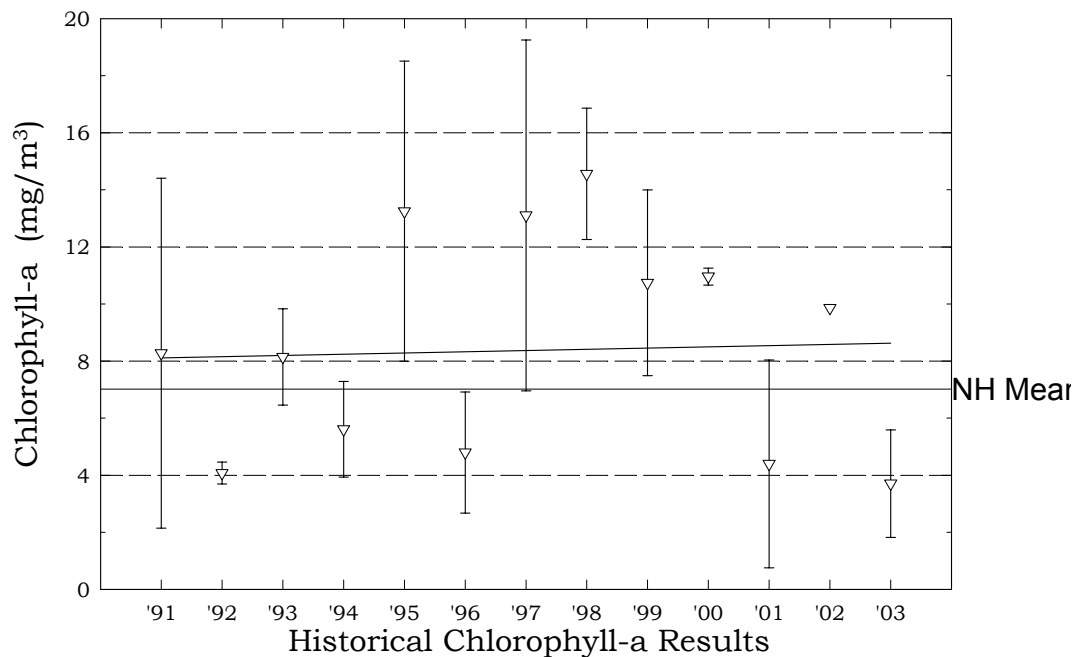
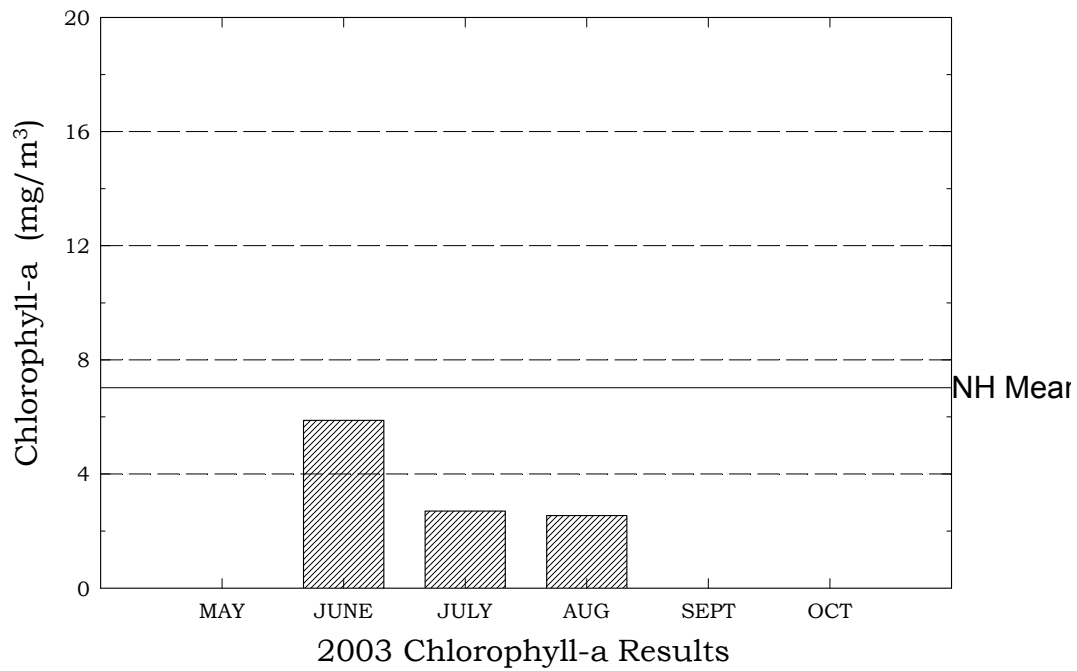
Proper Lawn Care In the Protected Shoreland, The Comprehensive Shoreland Protection Act, WD-SP-2, NHDES Fact Sheet, (603) 271-3503 or www.des.state.nh.us/factsheets/sp/sp-2.htm.

APPENDIX A

GPAPHS

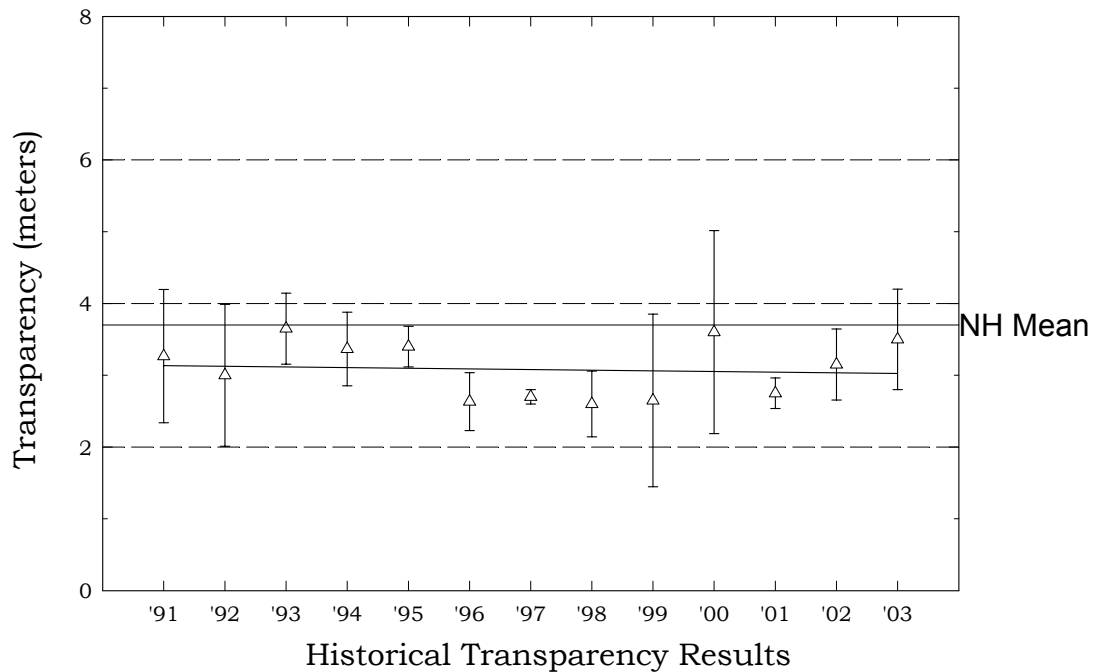
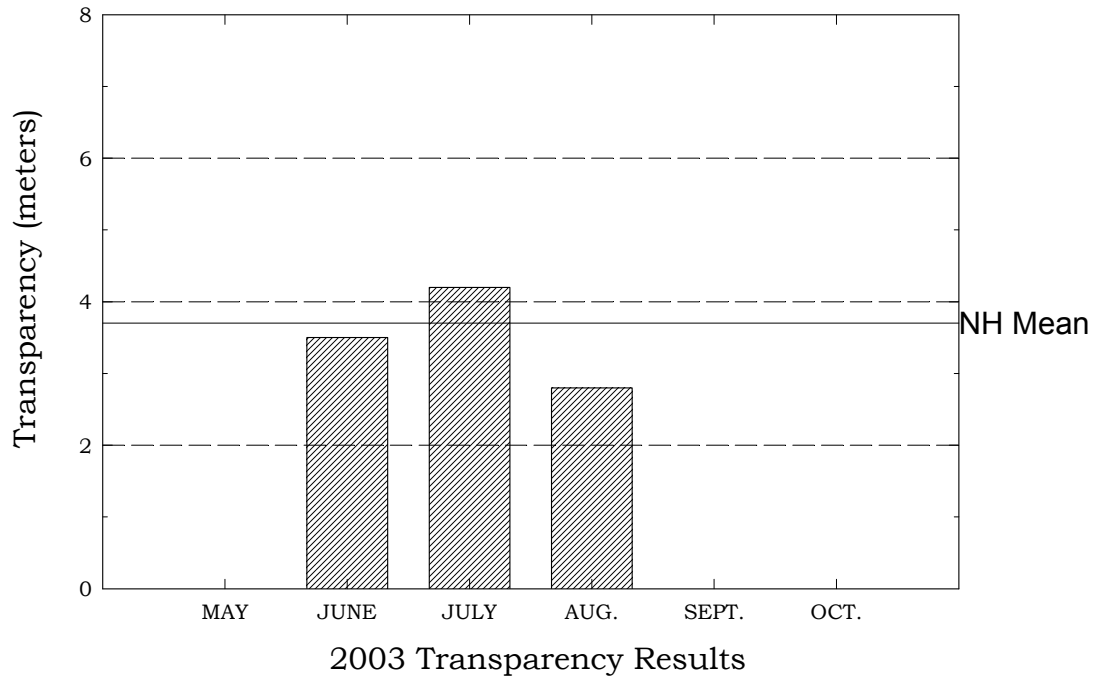
Forest Lake, Winchester

Figure 1. Monthly and Historical Chlorophyll-a Results



Forest Lake, Winchester

Figure 2. Monthly and Historical Transparency Results



Forest Lake, Winchester

Figure 3. Monthly and Historical Total Phosphorus Data.

